



REVIEW ARTICLE

IGF-1 biomarker testing in an ethical context

Annabelle Trojan¹, Beatriz H. Aristizabal², Lina M. Jay³, Tatiana Castillo⁴, Pedro J. Penagos⁵, Ignacio Briceño⁶, Jerzy Trojan^{4*}

¹ Faculty of Medicine, FUJNC – Corpas University, Bogota, Colombia

² Laboratory of Molecular Biology, HPTU Hospital and UNIGEM, Medellín, Colombia

³ Department of Investigation, INS – National Institute of Health, Bogota, Colombia

⁴ Department of Chemistry, UDFJC – Distrital University, Bogota, Colombia

⁵ Department of Neurosurgery, INC – National Institute of Cancerology, Bogota, Colombia

⁶ Institute of Human Genetics, PUJ – Javeriana University, Bogota, Colombia

Abstract: As we have come to know, there is a connection between cancer biomarkers and genes, along with their susceptibility to a particular disease, all of which have an obvious impact on the clinical practice and development of genetic testing. In any cancer disease, the diagnosis and treatment should be related to the investigation of specific biomarkers (generally antigens and proteins) and their corresponding genes. The study of different antigens such as alpha-fetoprotein, insulin-like growth factor I (IGF-I), insulin-like growth factor II, vascular endothelial growth factor, and epidermal growth factor, as well as their presence in neoplastic cells have demonstrated that IGF-I is an essential target for gene testing and therapeutic purpose. An over-expression of the IGF-I gene in mature tissues is a sign of neoplastic processes, *e.g.* brain or breast malignancy. A lot of questions have arisen regarding the ethics of gene testing, particularly concerns on the selection of patients for specific growth hormone/insulin-like growth factor I (GH/IGF-I) testing. Evidently, our current society is involved in a process of geneticization – the redefinition of individuals in terms of genetic codes. As such, we should take extreme care when making ethical judgments based on “scientific evidence” derived from genetic testing (typically those involving different biomarkers such as DNA, RNA, chromosomes, and proteins) in relation to genetic abnormalities that could predict current or future diseases. In this situation, the understanding of bioethics is of utmost importance.

Keywords: cancer; biomarkers; IGF-I; gene testing; bioethics

Citation: Trojan A, Aristizabal BH, Jay LM, Castillo T, Penagos PJ, *et al.* IGF-1 biomarker testing in an ethical context. *Adv Mod Oncol Res* 2016; 2(4): 188–200. <http://dx.doi.org/10.18282/amor.v2.i4.58>.

*Correspondence to: Jerzy Trojan, Department of Chemistry, Distrital University, Bogota, Colombia; genetherapy@hotmail.fr

Received: 04th October 2015; **Accepted:** 12th July 2016; **Published Online:** 15th August 2016

Introduction

After demonstrating the convergence between ontogenesis and oncogenesis using alpha-fetoprotein (AFP) as a new biomarker of neoplastic development^[1,2], the phenomenon was confirmed using another cancer biomarker—insulin-like growth factor I (IGF-I)^[3,4], precipitated by the development of IGF-I testing^[5,6] and the establishment of cancer gene therapy by applying the anti-IGF-I approach^[7]. Using the IGF-I biomarker, which

plays an important role in cancerology^[7] as an example, this review describes the common ethical problems faced by genetic testing.

A biomarker or molecular marker is defined by the National Cancer Institute (NCI) as “a biological molecule found in blood, other body fluids, or tissues (including RNA and microRNA) that is a sign of a normal or abnormal process, or of a condition or disease.” Biomarkers – especially those associated with genetic mutations or epigenetic alterations – help to identify early

stages of cancer, patient prognosis, treatment options, and response to therapy^[8-14]. New array-based technologies such as comparative genomic hybridization arrays (CGH), single nucleotide polymorphism (SNP) arrays, and protein arrays, among other things, are powerful tools when identifying biomarkers. In fact, cancer studies using this kind of technology have identified genes that are involved with the initiation, promotion, progression, and treatment response of cancer, as well as improved the understanding of the biological characteristics of cancer cells. Finally, changes in micro RNA (miRNA) expression can also be a biomarker, e.g. an increase in *miR-206* and *miR-221* gene expression or a down-expression of the *miR-125b* and *let-7* genes^[13,14].

As far as biomarkers are concerned, related genetic testing constitutes an important domain in clinical laboratory diagnostic. In this context, possible patents that are related to genetic testing should be discussed. The idea of patenting genes may seem absurd—as the “invention” is prior to the inventor, yet it is a reality. The United States, just 15 years ago, had at least 48 private companies with a minimum of three patents in class 435/6 (molecular biology involving nucleic acids)^[15]. As of today, the genes of plants, animals, and humans have been patented^[16,17]. While there is a scientific basis to patenting a method derived from the knowledge of one gene, it is contentious to patent a gene itself. If the human genome is (in a symbolic sense) a heritage of humanity (UNESCO, 1997)^[18], then it is common property. In addition, there is great concern that gene sequence patents may hinder future biotechnological innovations in the medical field^[19]. Under these circumstances, understanding bioethics is a priority. Jean Dausset, winner of the 1980 Nobel Prize of Medicine said in a personal correspondence with co-author Trojan J (unpublished, 2000): “Bioethics is an extremely important event in human consciousness. This stems from the extraordinary gap between concepts and technology due to the dazzling advances in biology and genetics.”

IGF-1 biomarker

Certain antigens, which behave as oncoproteins, are present in normal fetal/neonatal development but are absent from mature tissues. Among them, serum albumin, transferrin, AFP^[2], growth hormones such as epithelial growth factor (EGF), fibroblast growth factor (FGF), platelet-derived growth factor (PDGF), vascular endothelial growth factor (VEGF), transforming growth factor-beta (TGF-beta), and especially IGF I and II^[20,21] reappear in neoplastic developing tissues^[3,4,6,22] such as in brain, liver, prostate, ovarian, colon, and uterus can-

cers^[23-27].

Comparative studies of different antigens such as AFP, IGF-I, IGF-II, VEGF, and EGF present in neoplastic cells^[6,7,28] have demonstrated that IGF-I constitutes an important target for genetic testing and therapeutic purpose. The arrest of IGF-I expression diminishes or stops neoplastic development^[7,29,30] due to the consequential apoptosis and anti-tumor immune response (MHC-I)^[31-33]. These two phenomena, which play an important role in the mechanism of IGF-I, are barely present (if not non-existent) in other oncoprotein and growth factor mechanisms such as EGF, VEGF, or TGF-beta^[34-37]. In terms of the growth factors involved in both ontogenesis and carcinogenesis, IGF-I may be a highly promising therapeutic and diagnostic target (400 publications/year). Similar to AFP, IGF-I is involved in tissue development and differentiation, especially in the development of the nervous system^[1,38,39] as a mediator of growth hormone, thyroid stimulating hormone (TSH), and glucose metabolism, acting locally with autocrine/paracrine and has a predominant role compared to other growth factors^[20,39-45].

According to Baserga^[46], IGF-I is one of the most important growth factors that is related to normal and neoplastic differentiation, and its overproduction is considered to be a participating factor in cancer development^[33,44,47,48]. IGF-I has been reported to block a number of apoptosis pathways (IRS/PI3K/AKT/Bcl, GSK3, Ca²⁺, or caspases)^[46,49-54]. Moreover, the mechanism of IGF-I depends on its receptor – IGF-I-R, which plays a predominant role in tumor growth processes^[43,47-49,55]. IGF-I constitutes the first step of the following signal transduction pathway: IRS/PI3K-PKC/PDK1/AKT-Bcl2/GSK3/GS^[56,57]. The elements of the aforementioned IGF-I-related transduction pathway were also considered as targets for diagnostic and therapeutic purposes^[37,39,54,56,58-68]. The measurement of IGF-I and insulin-like growth factor-binding protein (IGFBP-3) often serves as first-line testing in children with growth disorders. The role of acid-labile subunit (ALS) as a screening parameter for homozygous or heterozygous mutations of the *ALS* gene has been recently determined^[69]. The relationship between IGF-I and IGF binding proteins are being introduced in clinical diagnostics as one of the indicators of precancerous development^[70]. The serum level of IGF-I (considered as a marker) was introduced in the diagnostic of breast cancer^[71-73], prostate cancer^[74,75], colorectal cancer^[76-78], lung cancer^[79-80], and pancreatic cancer^[77,81].

The substantial individual variability in the circulating levels of IGF-I and binding proteins (especially IGFBP-3) may be important in determining the risk of developing

malignant prostate, breast, colorectal, lung, and liver tumors^[82-84]. Since the introduction of IGF-I in the 1990s as a breast cancer biomarker, another new biomarker – mammaglobin, a protein that is a member of the globin secreting family and contains lypophilin B was recently proposed for clinical diagnostic^[14]. As far as the relationship between cancer and depression is concerned, elevated IGF-I serum levels have been found to be significantly associated with depression. This suggests that IGF-I signaling could play a role in the pathophysiology of depression and could possibly influence the response to antidepressant treatment^[85].

Genetic engineering

Molecular biologists consider that the processes of living beings adhere to the laws of physics, chemistry, and protein chemistry^[1,86-88]. However, life is characterized by high variability; especially when it comes to brain development, as demonstrated by the murine model of neoplastic central nervous system (CNS) development^[22]. Major advances in molecular biology have led to the sequence hypothesis, from nucleotides to amino acids, which is consistent but independent of the laws of physics and chemistry^[89]. Yet, should our fate be sealed by our genes and reduced to physics and chemistry? We should therefore take extreme care in making or accepting ethical judgments based on “scientific evidence”^[90].

Molecular biology has allowed genetics to evolve a structure-function relationship. These findings generate deep questions about how biological structures operate, manage, and affect evolution^[72,78,88,91,92]. Although molecular biology must always be consistent with physical and chemical processes, it cannot be derived from physics and chemistry alone^[88,93]. Our knowledge of molecular and cellular biology techniques such as recombinant DNA and cloning has led to the development of a new domain – biotechnology. Biotechnology includes methods ranging from genetic engineering and genetic mapping to tissue culture hybridomas and genetically engineered vaccine. Genetic engineering has allowed the isolation and manipulation of genes to take place. Its impact has been particularly important as it has led to the creation of transgenic animals used as experimental models for research and the observation of genetic diseases, and to the creation of organic substances for therapeutic purposes through the process of enabling or disabling of a gene^[6,27,94-97].

The creation in the early 1990s of a new medical domain, termed gene therapy, has become the most im-

portant revolution in the treatment of different diseases such as cancers, infections, and fetal diseases. Gene therapy is the logical consequence of genetic testing as both target the same gene and its related protein. The first gene therapy case approved in the United States took place in 1990 at the National Institute of Health (NIH)^[98]. W. F. Anderson developed a treatment for his patient with a genetic defect that resulted in adenosine deaminase deficiency-severe combined immune deficiency (ADA-SCID). The effects were only temporary but successful nonetheless. In the same regard, the first stem cell gene therapy by C. Bordignon *et al.* was performed in 1992 at the Vita-Salute San Raffaele University, Milan, Italy, using hematopoietic stem cells as vectors to deliver genes intended to correct hereditary diseases^[99]. Clinical trials resumed following regulatory review of the protocol in the United States, the United Kingdom, France, Italy, and Germany.

The first cancer gene therapy was introduced in 1992/93 at Case Western Reserve University (CWRU), Cleveland using the IGF-I antisense approach for the treatment of glioblastoma multiforme (GBM), the most common of human brain tumor whose outcome is always fatal^[7] and the protocol was approved by NIH in 1993 and FDA in 1994. The modified strategy, using anti-gene (antisense/triple helix) anti IGF-I technology, has shown promising results in clinical trials; the median survival of glioblastoma patients reached 21 months and in some cases, reaching up to three to four years. This strategy was also proven to be efficient in the treatment of six different cancer diseases (as reported by the NATO Science program on cancer gene therapy: USA, France, Poland, Germany; 2002–2007)^[27,39].

To target specific genetic defects, different kinds of molecules (antibodies, antisense oligodeoxynucleotides, antisense cDNAs, short peptides, and other small molecules) have been employed. The antisense technology has become one of the important anti-cancer approaches used in the last 10 years in preclinical and clinical studies of tumors, including GBM^[39,68,100]. However, genetic engineering presents a certain level of risk to the environment, plant and animal species, and especially mankind owing to the lack of knowledge concerning the effects induced by genetic manipulations in organisms. For this reason, NIH and the Food and Drug Administration (FDA) generally avoid gene therapy protocols that propose adenoviral or retroviral vectors, suggesting instead the use of episomal vectors, which present a small risk of incorporating DNA derived from genetic engineering manipulation into the human genome^[101].

Genetic testing

Genetic testing involves analyzing DNA, RNA, chromosomes, proteins, and metabolite abnormalities that could predict current or future diseases^[102,103]. In medicine, there is a tendency of using a genetic model to explain a particular disease, thus increasing the influence of genetic technologies in clinical practice^[104]. The criteria which determines “who should be tested” depends on the type of disease; however, it is recommended that test subjects have a family history of at least three generations^[105]. This effectively identifies high-risk individuals who would benefit from genetic testing and appropriate prophylactic measures, as well as early therapy, according to the 1996 report from the American Society of Clinical Oncology^[106].

Genetic testing involves analyzing detailed family history, determining the type of test, and interpreting the results. Knowledge on available treatments and preventive measures is important for genetic counseling, which is an essential component in ensuring adequate data collection of family history, risks, and the selection of appropriate tests. Genetic counseling should be offered to all patients before and after genetic testing. Genetic tests have psycho-social implications owing to the risk of improper handling of information by insurers, as well as the loss of privacy and the potential to generate anxiety and/or depression in the patient and their family^[107,108]. The use of genetic testing has improved the survival rate of people at risk. Advances in genetics and genetic testing are increasing rapidly, hence implying a greater responsibility in the management of patients undergoing predictive or diagnostic tests. As genetic testing involves patients and their family members, clinicians must be well-informed of the medical indications for genetic testing and the different available examinations, as well as having the ability to analyze and interpret the results.

IGF-1 gene test

There is a connection between genes and their suscepti-

bility to certain diseases. Certain genes are related to different pathologies such as cancer, diabetes, hypertension, and obesity, among other things^[109]. Some of these data have an impact on clinical practice, generating the availability of genetic tests. For example, knowledge of the genes responsible for colorectal cancer has resulted in improved genetic testing, management, and early treatment of the disease^[110]. As far as the gene-disease relationship is concerned, we need to think of genes as structures that induce protein synthesis and therefore, they are potential tissue markers. As far as the *IGF-1* gene is concerned, an over-expression of the *IGF-1* gene in mature tissues is a sign of neoplastic processes, especially brain tumors^[111] (Figure 1), and also a sign of other neural pathologies such as Huntington disease^[111] (unpublished data) (Figure 2) or depression^[112]. On the contrary, the deletion of the *IGF-1* gene is associated with reduced brain growth and mental retardation^[42,113].

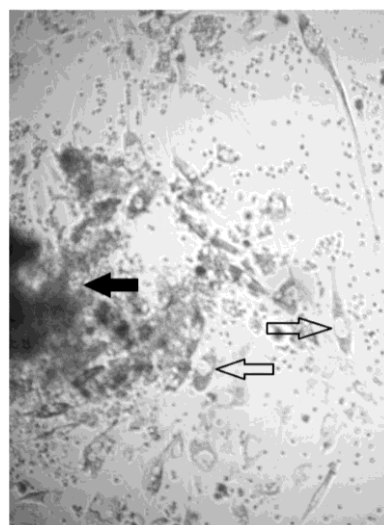


Figure 1. *In vitro* staining of IGF-1 biomarker in glioma cell culture; sixth day of culture established from human glioblastoma biopsy. Note the cells (empty arrows) proliferating from compact tissue of biopsy (black arrow). The tissue and cells are stained for IGF-1 using anti IGF-1 antibodies applied in immunoperoxidase technique (note the dark cytoplasm)^[1].

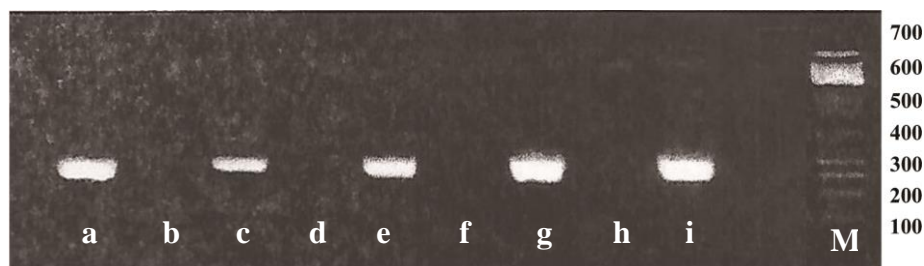


Figure 2. IGF-1 genetic testing in nervous system diseases using PCR technique^[118]. Solid tumor of glioblastoma (a); Blood samples removed from glioblastoma patients (c, e); Blood samples removed from patients with Huntington disease^[119] (g, i); Blood samples removed from healthy patients (b, d, f, h); Marker (M): PCR markers, Promega Corporation. Note the 200–300 bp of positive bands (a, c, e, g, i).

Molecular testing could be useful in congenital malformations involving the central nervous system, which continue to be a major cause of infant death in the Western world; the incidence of malformations being higher in children with intrauterine growth retardation. Primary malformations go hand-in-hand with genetic intrinsic diseases^[114], and the increase of intracytoplasmic IGF-I is associated with CNS malformations^[115]. IGF-I function is parallel to the commonly used AFP marker, which becomes useful in the molecular diagnostics of neonatal malformations and tumor diseases^[116]. These observations have enabled the testing of IGF-I as an oncoprotein and genetic marker. As a result of the “IGFs and Cancer” Symposium (held in Halle, Germany; September 15–17, 2000), an increased IGF-I serum level and an increased *IGF-I* gene expression in mature tissues have been viewed as a putative diagnostic marker for biological activity in different tumors as 17 different tumors are believed to express the *IGF-I* gene^[82].

There is a convergence between normal embryo/fetal development and neoplastic development, more specifically the neoplastic brain development^[1,117]. According to the theory of evolution, life is derived from amino acids; therefore, life can be altered by the imbalance that is related to protein presence. Diagnosis and treatment should logically be related to the investigation of proteins or growth factors (specific antigens) and their corresponding genes; firstly, by using gene testing for diagnosis^[5,118,119] (Figure 2) and subsequently, targeting specific genes through special therapy such as cancer gene therapy^[7,120] (Figure 3). Two promoters control IGF-I expression^[42,121], with a low serum level of IGF-I being related to the IGF-I first promoter activity (*i.e.*, nucleotide sequence changes), as demonstrated in children with growth disorders presenting normal level of growth hormone (GH)^[118]. The last data showed that testing the IGF-I first promoter region using polymerase chain reaction/single-strand conformation polymorphism (PCR/SSCP) analysis could be useful in the diagnosis of

growth disorders^[118].

In another study of genetic screening, which looks into the criteria of short children, the genetic analysis of these children with normal birth size has led to the detection of a *SHOX* or *IGF1R* genetic variant in 6% of short children^[122]. According to Wit^[123], if no obvious candidate gene can be determined in short children's genetic testing, a whole genome approach can be taken in order to check for deletions, duplications and/or uniparental disomy, or whole exome sequencing. Curiously, IGF-I plays also a role in the control of eye growth; IGF-I polymorphisms are associated with myopia (IGF-I genotyping was performed with selected tag single nucleotide polymorphisms)^[124]. In recent studies regarding prostate cancer survival, IGF-I pathway genetic polymorphisms, in parallel with the circulating levels of IGF-I and IGFBP-3, have demonstrated that *IGF2-AS* and somatostatin receptor 2 (*SSTR2*) genes are primarily associated with pancreatic cancer mortality. Therefore, the testing of these two genes may be important in determining the survival of pancreatic cancer patients^[125]. Moreover, the genetic variation in the *IGF-I*, *IGFBP-3*, and *SSTR-2* genes (wherein SNPs were genotyped) seems to influence the circulating levels of IGF-I and IGFBP-3 in prostate and breast cancers^[126].

Moreover, genetic association and sequencing of the insulin-like growth factor 1 gene in bipolar disorder patients (via haplotype association and a gene test with wide significance of permutation testing for all markers genotyped IGF-1) implicate *IGF-1* as a candidate gene that causes genetic susceptibility to this psychiatric disease^[112]. The study of IGF-I genetic variation in GH/IGF-1/insulin signaling pathway has demonstrated a potentially new human longevity loci^[127]. We need to underline that there is a strong relation between genetics, signal transduction pathway, and metabolism. Gene coding for IGF-I and other growth factor-induced signaling, in particular the PI3K/AKT/mTOR pathway and in relation with rapamycin, promotes anabolism and suppresses

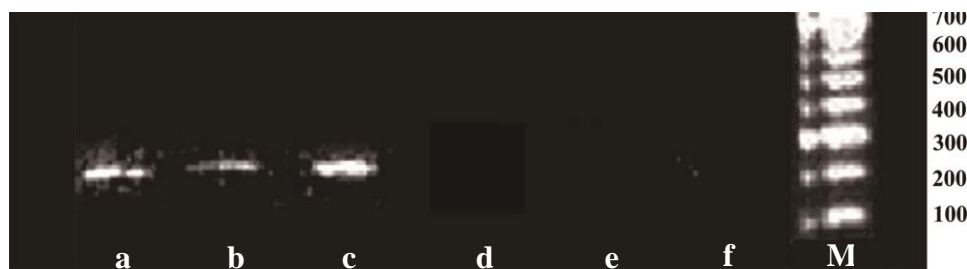


Figure 3. IGF-1 gene testing in cancer diseases. RT-PCR technique^[120]. Parental cultured cells derived from (a) glioblastoma, (b) hepatocarcinoma, (c) ovary cyst-adenocarcinoma. Absence of *IGF-1* expression (d, e, f) in the same cells transfected with antisense anti IGF-1 vector (cell “vaccines”). (M): Marker. Note the 200–300 bp positive bands (a, b, c). The parental and transfected cells illustrate the passage from gene testing to gene therapy, respectively.

catabolism to produce energy and macromolecules, respectively^[128]. The interruption of any of these metabolic effects renders the growth factor ineffective.

Cancer is a prime example of a common human disease with genetically-defined, pathological metabolic perturbations^[129-131]. For cancer, enzyme mutations may function as oncogenes. Genomic sequencing of tumors expressing IGF-I, especially gliomas^[132,133], has identified mutations in two isoforms of NADP⁺-dependent isocitrate dehydrogenase (IDH1 and IDH2). Tumor tissue and cell lines expressing mutant IDH1 or IDH2 produce large quantities of the (D)-2-hydroxyglutarate (2-HG) metabolite. This metabolite is produced from the NADPH-dependent reduction of α -ketoglutarate to 2-HG. The presence of 2-HG in these tumors mirrors one of the best established connections between inborn errors of metabolism (IEM) and cancer.

Bioethics

As far as bioethics and gene testing are concerned, the connection between genes and their susceptibility to certain diseases should be discussed. An increase in *IGF-I* gene expression has been viewed as a “sign” of potential tumor development in tested patients^[70,82-84]. The question is: Should this type of test (which does not guarantee the appearance of cancer) be communicated to patients, relatives, or insurance companies? Another example of ethical issues for parents and physicians is the case of genetic testing in short children: Who decides if genetic testing is appropriate for this type of “pathology”?^[134]. These ethical questions also concern patients who are selected for specific genetic testing of the GHIIGF-I axis, based on previously obtained clinical and biochemical assessments of growth deficiencies^[135].

On November 11, 1997, the United Nations’ Educational, Scientific, and Cultural Organization (UNESCO) adopted the Universal Declaration on the Human Genome and Human Rights. The text refers to the need of educating the society on bioethics and institutionalizes the presence of bioethics committees in the decision making process. Thus, the 186 countries involved in UNESCO recognized the need to: (a) promote education in bioethics, at all levels; (b) let individuals and society know of their collective responsibility in defending human dignity in topics related to biology, genetics, and medicine; (c) encourage open social and international debate, as well as ensuring freedom of expression involving the different currents of thought, should it be socio-cultural, religious, or philosophical; and (d) promote the creation (at the appropriate levels) of independent interdisciplinary and pluralistic bioethics com-

mittees. With respect to the function of the different committees, bioethical issues such as human genetic manipulation (human DNA, cells, individuals, and populations), human reproduction and embryology (the human embryo as the beginning of life and individualization, as well as assisted reproduction, embryo research and cloning), and genetically modified organisms (microorganisms released into the environment, with potential to evolve into transgenic animals and plants) should be handled appropriately.

Genetic testing comes with benefits as well as limitations. Individuals with “normal” genetic test results could experience relief whereas “abnormal” results may affect not only the patient, but also their family members. Informed carriers do benefit from knowing the risks associated with their disease, yet could still experience anxiety and guilt owing to possible transmission to the next generation^[136], along with the loss of privacy, and genetic discrimination from insurers and employers^[137,138]. Several insurance companies use the results of genetic testing in prenatal care to formulate insurance contracts, as well as to implement new policies and to derive concepts of health and disease, disorder, and abnormality^[104]. Genetic testing can potentially impose a bias on human beings, conveniently forgetting that every person including newborns are protected by the law, as stated in Article 6 of the Universal Declaration of Human Rights (UDHR): “Everyone has the right to recognition everywhere as a person before the law.”

The field of bioethics has evolved dramatically in recent years, and several major developments have transpired with regard to the niche area of biomedical sciences^[139,140]. It would be a terrible mistake to think of genes as “genes of a particular factor”^[141]. Perhaps the best example of “genes of a particular factor” is the gene of intelligence^[142], the gene of schizophrenia^[143], the gene of homosexuality^[144], and the gene of a particular behavior, among other things. The growing impact of genetic concepts in popular culture has been linked to “genetic essentialism”, the belief that human beings in all their complexity are products of a molecular model^[145]. To identify and analyze the cultural processes involved in biomolecular life sciences, it is important to clarify the concept of “geneticization”.

The concept of “geneticization” tries to describe the mechanisms of interaction between medicine, genetics, society, and culture^[146]. We can define geneticization as the socio-cultural interpretation and explanation of human beings using the terminology and concepts of genetics; a process of not only seeing health and disease as what they are, but also to observe all human behaviors and social interactions through the prism of biomolecular

technology^[147]. Genetic technology should not be regarded simply as a new technology that is available to enrich the knowledge of responsible autonomous consumers, but also as a tool capable of transforming our understanding of human existence. More than a field of science, genetics is a way of thinking – an ideology where “genetics is the answer”^[148].

Our society is clearly involved in the process of geneticization^[147]. This process involves a redefinition of individuals in terms of genetic codes (DNA). Disease, health, and body are explained in terms of molecular biology^[145]. It seems that the meaning of DNA is similar to that of the “immortal soul” as described in Christian theology. An example of the process of geneticization is the research programs and genetic counseling for β -thalassemia patients^[104]. Case in point, individuals in Cyprus may only marry if they have a certificate proving their participation in genetic research. The discussion on bioethics of geneticization should involve a moral dimension. The concept of geneticization can actually produce a change in focus; we can direct the attention of our society to the different dimensions of genetic technology, which is usually neglected in bioethical analysis.

Conclusion

Progress in the field of molecular biology, medicine, and related disciplines has changed our perception of life and death, and influenced our bioethical decisions. Through the knowledge of the human genome, one can easily see the body as a machine made of multiple interchangeable parts^[149]. The danger is that simplistic models of the body can override the science of life, interfering with its most sophisticated and complex mechanisms^[150]. Molecular advances have extended the possibilities of genetic testing, establishing a new category of “potential patients”. What is needed here is a redefinition of the concept of disease, focusing not only on clinical symptoms and genetic abnormality, but also on the increased risk of adverse individual consequences^[151].

The rapid advances in genetics have had an undeniable impact on society; the consensus, therefore, is that bioethics and biomedicine must always be accompanied by bio-criticism. Bioethics is no longer an issue debated only in developed countries or an issue dealt exclusively by large corporations^[120]. A number of discoveries that could revolutionize the economy and change our way of life^[7,94,119,152] have repercussions on new forms of diagnosis (*e.g.*, genetic testing) and therapies (*e.g.*, gene therapy), with the latter being associated with ethical considerations. In fact, although somatic

gene therapy is included in current clinical treatment, fetal gene therapy remains a subject of ethical interrogations in Western medicine.

Author contributions

All authors, in accordance to their respective specialties, have contributed to the preparation and final corrections of this manuscript.

Conflict of interest

The authors declare no potential conflict of interest with respect to the research, authorship, and/or publication of this article.

References

1. Trojan J, Uriel J, Deugnier MA, Gaillard J. Immunocytochemical quantitative study of alpha-fetoprotein in normal and neoplastic neural development. *Dev Neurosci* 1983–1984; 6(4–5): 251–259. doi: 10.1159/000112352.
2. Trojan J, Uriel J. (French) [Intracellular localisation of alpha-fetoprotein and serum albumin in the central nervous system of the rat during fetal and postnatal development]. *CR Acad Sci Paris* 1979; 289(15): 1157–1160.
3. Sandberg AC, Engberg C, Lake M, von Holst H, Sara VR. The expression of insulin-like growth factor I and insulin-like growth factor II genes in the human fetal and adult brain and in glioma. *Neurosci Lett* 1988; 93(1): 114–119. doi: 10.1016/0304-3940(88)90022-5.
4. Kiess W, Lee L, Graham DE, Greenstein L, Tseng LYH, *et al.* Rat C6 glial cells synthesize insulin-like growth factor I (IGF-I) and express IGF-I receptors and IGF-II/mannose 6-phosphate receptors. *Endocrinol* 1989; 124(4): 1727–1736. doi: 10.1210/endo-124-4-1727.
5. Johnson TR, Trojan J, Rudin SD, Blossey BK, Ilan J, *et al.* Effects of actinomycin D and cycloheximide on transcript levels of IGF-I, actin, and albumin in hepatocyte primary cultures treated with growth hormone and insulin. *Mol Reprod Dev* 1991; 30(2): 95–99. doi: 10.1002/mrd.1080300204.
6. Trojan J, Blossey BK, Johnson TR, Rudin SD, Tykocinski M, *et al.* Loss of tumorigenicity of rat glioblastoma directed by episome-based antisense cDNA transcription of insulin-like growth factor I. *Proc Natl Acad Sci USA* 1992; 89(11): 4874–4878. doi: 10.1073/pnas.89.11.4874.
7. Trojan J, Johnson TR, Rudin SD, Ilan J, Tykocinski ML, *et al.* Treatment and prevention of rat glioblastoma by immunogenic C6 cells expressing antisense insulin-like growth factor I RNA. *Science* 1993; 259(5091): 94–97. doi: 10.1126/science.8418502.
8. NCI Dictionary of Cancer Terms [Internet]. National Cancer Institute (US); 2015 [cited 2016]. Available from:

- <http://www.cancer.gov/publications/dictionaries/cancer-terms>.
9. Trojan J, Naval X, Johnson T, Lafarge-Frayssinet C, Hajeri-Germond M, et al. Expression of serum albumin and of alphafetoprotein in murine normal and neoplastic primitive embryonic structures of teratocarcinoma. *Molec Reprod Dev* 1995; 42 (4), 369–378. doi: 10.1002/mrd.1080420402
 10. Hu B, Niu X, Cheng L, Yang LN, Li Q, et al. Discovering cancer biomarkers from clinical samples by protein microarrays. *Proteomics Clin Appl* 2015; 9(1–2): 98–110. doi: 10.1002/prca.201400094.
 11. Trojan LA, Kopinski P, Wei MX, Ly A, Glogowska A, et al. IGF-I: From diagnostic to triple-helix gene therapy of solid tumors. *Acta Biochim Pol* 2002; 49(4): 979–990.
 12. Verma M and Manne U. Genetic and epigenetic biomarkers in cancer diagnosis and identifying high risk populations. *Critical Rev Oncol Hematol* 2006; 60(1): 9–18. doi: 10.1016/j.critrevonc.2006.04.002.
 13. Cruz Tapias PA, Villegas Gálvez VE, Ramírez Clavijo SR. Fundamento biológico y aplicación clínica de los marcadores tumorales séricos (Spanish) [Biological basis and clinical application of serum tumor markers]. *Rev Cienc Salud* 2008; 6(2): 85–98.
 14. Galvis-Jiménez JM, Curtidor H, Patarroyo MA, Monterrey P, Ramírez Clavijo SR, et al. Mammaglobin peptide as a novel biomarker for breast cancer detection. *Cancer Biol Ther* 2013; 14(4): 327–332. doi: 10.4161/cbt.23614.
 15. Henry MR, Cho MK, Weaver MA, Merz JF. DNA patenting and licensing. *Science* 2002; 297(5585): 1279. doi: 10.1126/science.1070899.
 16. Baird P. Patenting and human genes. *Perspect Biol Med* 1998; 41(3): 391–408. doi: 10.1353/pbm.1998.0061.
 17. Poland SC. Genes, patents, and bioethics—Will history repeat itself? *Kennedy Inst Ethics J* 2000; 10(3): 265–281. doi: 10.1353/ken.2000.0022.
 18. Byk C. A map to a new treasure island: The human genome and the concept of common heritage. *J Med Philos* 1998; 23(3): 234–246. doi: 10.1076/jmep.23.3.234.2589.
 19. Heller MA, Eisenberg RS. Can patents deter innovation? The anticommens in biomedical research. *Science* 1998; 280(5364): 698–701. doi: 10.1126/science.280.5364.698.
 20. Froesch ER, Schmid C, Schwander J, Zapf J. Actions of insulin-like growth factors. *Annu Rev Physiol* 1985; 47: 443–467. doi: 10.1146/annurev.ph.47.030185.002303.
 21. Han VK, D’Ercole AJ, Lund PK. Cellular localization of somatomedin (Insulin-like growth factor) messenger RNA in the human fetus. *Science* 1987; 236(4798): 193–197. doi: 10.1126/science.3563497.
 22. Trojan J, Uriel J. Localisation of alphafetoprotein (AFP) in murine teratocarcinoma. *Biomedicine* 1981; 34(3): 140–146.
 23. Chatel M, Bourg V. (French) [Intracerebral tumours]. *Rev Prat* 2004; 54(8): 889–896.
 24. Guha A, Mukherjee J. Advances in the biology of astrocytomas. *Curr Opin Neurol* 2004; 17(6): 655–662. doi: 10.1097/00019052-200412000-00004.
 25. Ohgaki H, Dessen P, Jourde B, Horstmann S, Nishikawa T, et al. Genetic pathways to glioblastoma: A population-based study. *Cancer Res* 2004; 64(19): 6892–6899. doi: 10.1158/0008-5472.CAN-04-1337.
 26. Wrensch M, Rice T, Miike R, McMillan A, Lamborn KR, et al. Diagnostic, treatment, and demographic factors influencing survival in a population-based study of adult glioma patients in the San Francisco Bay Area. *Neuro Oncol* 2006; 8(1): 12–26. doi: 10.1215/S1522851705000268.
 27. Trojan J, Pan YX, Wei MX, Ly A, Shevelev A, et al. Methodology for anti-gene anti-IGF-I therapy of malignant tumours. *Chemother Res Pract* 2012(2012). doi: 10.1155/2012/721873.
 28. Hajeri-Germond M, Naval J, Trojan J, Uriel J. The uptake of alpha-foetoprotein by C-1300 mouse neuroblastoma cells. *Br J Cancer* 1985; 51: 791–797. doi: 10.1038/bjc.1985.123.
 29. Lafarge-Frayssinet C, Duc HT, Sarasin A, Anthony D, Guo Y, et al. Antisense IGF-I transferred into a rat hepatoma cell line inhibits tumorigenesis by modulating MHC-I cell surface expression. *Cancer Gene Ther* 1997; 4(5): 276–285.
 30. Anthony DD, Pan YX, Wu SG, Shen F, Guo J. *Ex vivo* and *in vivo* IGF-I antisense RNA strategies for treatment of cancer in humans. In: Walden P, Trefzer U, Sterry W, Farzaneh F, Zambon P, (editors). *Gene therapy of cancer*. New York: Springer US; 1998. p. 27–34. doi: 10.1007/978-1-4615-5357-1_5.
 31. Trojan J, Duc HT, Upegui-Gonzalez LC, Hor F, Guo Y, et al. Presence of MHC-I and B-7 molecules in rat and human glioma cells expressing antisense IGF-I mRNA. *Neurosci Lett* 1996; 212(1): 9–12. doi: 10.1016/0304-3940(96)12770-1.
 32. Upegui-Gonzalez LC, Duc HT, Buisson Y, Arborio M, Lafarge-Frayssinet C, et al. Use of the IGF-I antisense strategy in the treatment of the hepatocarcinoma. In: Walden P, Trefzer U, Sterry W, Farzaneh F, Zambon P, (editors). *Gene therapy of cancer*. New York: Springer US; 1998. p. 35–42. doi: 10.1007/978-1-4615-5357-1_6.
 33. Kooijman R. Regulation of apoptosis by insulin-like growth factor (IGF)-I. *Cytokine Growth Factor Rev* 2006; 17(4): 305–323. doi: 10.1016/j.cytogfr.2006.02.002.
 34. Riedel F, Götte K, Li M, Hörmann K, Grandis JR. Abrogation of VEGF expression in human head and neck squamous cell carcinoma decreases angiogenic activity *in vitro* and *in vivo*. *Int J Oncol* 2003; 23(3): 577–583. doi: 10.3892/ijo.23.3.577.
 35. Reardon DA, Quinn JA, Vredenburgh JJ, Gururangan S, Friedman AH, et al. Phase 1 trial of gefitinib plus siroli-

- mus in adults with recurrent malignant glioma. *Clin Cancer Res* 2006; 12(3): 860–868. doi: 10.1158/1078-0432.CCR-05-2215.
36. Lassman AB, Rossi MR, Razier JR, Abrey LE, Lieberman FS, *et al.* Molecular study of malignant gliomas treated with epidermal growth factor receptor inhibitors: Tissue analysis from North American Brain Tumor Consortium Trials 01-03 and 00-01. *Clin Cancer Res* 2005; 11(21): 7841–7850. doi: 10.1158/1078-0432.CCR-05-0421.
 37. Schlingensiepen KH, Jaschinski F, Lang SA, Moser C, Geissler EK, *et al.* Transforming growth factor-beta 2 gene silencing with trabedersen (AP 12009) in pancreatic cancer. *Cancer Sci* 2011; 102(6): 1193–1200. doi: 10.1111/j.1349-7006.2011.01917.x.
 38. Ostos H, Astaiza G, Garcia F, Bautista M, Rojas F. Disminución de la incidencia de defectos de cierre del tubo neural en el Hospital Universitario de Neiva: Posible efecto de la promoción del consumo de ácido fólico (Spanish) [Decreased incidence of defects of neural tube closure at the University Hospital of Neiva: Possible effect promotion of folic acid]. *Biomédica* 2000; 20(1): 18–24. doi: 10.7705/biomedica.v20i1.1043.
 39. Trojan J, Cloix JF, Ardourel MY, Chatel M, Anthony DD. Insulin-like growth factor type I biology and targeting in malignant gliomas. *Neurosci* 2007; 145(3): 795–811. doi: 10.1016/j.neuroscience.2007.01.021.
 40. Le Roith D, Bondy C, Yakar S, Liu JL, Butler A. The somatomedin hypothesis: 2001. *Endocr Rev* 2011; 22(1): 53–74. doi: 10.1210/edrv.22.1.0419.
 41. Bermudez AJ, Salinas S. Recomendación sobre el uso de pruebas rápidas para TSH neonatal (Spanish) [Recommendation on the use of rapid tests for neonatal TSH]. *Informe Quincenal-Epidemiol Nac* 2002; 7(7): 102–106.
 42. Le Roith D. The insulin-like growth factor system. *Exp Diabetes Res* 2003; 4(4): 205–212. doi: 10.1155/EDR.2003.205.
 43. Baserga R. The insulin-like growth factor-I receptor as a target for cancer therapy. *Expert Opin Ther Targets* 2005; 9(4): 753–768. doi: 10.1517/14728222.9.4.753.
 44. Adhami VM, Afaq F, Mukhtar H. Insulin-like growth factor-I axis as a pathway for cancer chemoprevention. *Clin Cancer Res* 2006; 12(19): 5611–5614. doi: 10.1158/1078-0432.CCR-06-1564.
 45. Chen H, Mester T, Raychaudhuri N, Kauh, CY, Gupta S, *et al.* Teprotumumab, an IGF-1R blocking monoclonal antibody inhibits TSH and IGF-1 action in fibrocytes. *J Clin Endocrinol Metab* 2014; 99(9): E1635–1640. doi: 10.1210/jc.2014-1580.
 46. Baserga R. Oncogenes and the strategy of growth factors. *Cell* 1994; 79(6): 927–930. doi: 10.1016/0092-8674(94)90023-X.
 47. Pollak MN, Schernhammer ES, Hankinson SE. Insulin-like growth factors and neoplasia. *Nat Rev Cancer* 2004; 4: 505–518. doi:10.1038/nrc1387.
 48. Kurmasheva RT, Houghton PJ. IGF-I mediated survival pathways in normal and malignant cells. *Biochim Biophys Acta* 2006; 1766(1): 1–22. doi:10.1016/j.bbcan.2006.05.003.
 49. Baserga R, Sell C, Porcu P, Rubini M. The role of the IGF-I receptor in the growth and transformation of mammalian cells. *Cell Prolif* 1994; 27(2): 63–71. doi: 10.1111/j.1365-2184.1994.tb01406.x.
 50. Delaney CL, Cheng HL, Feldman EL. Insulin-like growth factor-I prevents caspase-mediated apoptosis in Schwann cells. *J Neurobiol* 1999; 41(4): 540–548. doi: 10.1002/(SICI)1097-4695(199912)41:4<540::AID-NEU9>3.0.CO;2-P.
 51. Crowder RJ, Freeman RS. Glycogen synthase kinase-3 β activity is critical for neuronal death caused by inhibiting phosphatidylinositol 3-kinase or Akt but not for death caused by nerve growth factor withdrawal. *J Biol Chem* 2000; 275(44): 34266–34271. doi: 10.1074/jbc.M006160200.
 52. Mason JL, Ye P, Suzuki K, D’Ercole AJ, Matsushima GK. Insulin-like growth factor-1 inhibits mature oligodendrocyte apoptosis during primary demyelination. *J Neurosci* 2000; 20(15): 5703–5708.
 53. Chrysis D, Calikoglu AS, Ye P, D’Ercole AJ. Insulin-like growth factor-I overexpression attenuates cerebellar apoptosis by altering the expression of Bcl family proteins in a developmentally specific manner. *J Neurosci* 2001; 21(5): 1481–1489.
 54. Trojan J, Anthony DD. Antisense strategies in therapy of gliomas. *Curr Signal Transduct Ther* 2011; 6(3): 411–423. doi: 10.2174/157436211797483895.
 55. Camirand A, Pollak M. Co-targeting IGF-1R and c-kit: Synergistic inhibition of proliferation and induction of apoptosis in H 209 small cell lung cancer cells. *Br J Cancer* 2004; 90(9): 1825–1829. doi: 10.1038/sj.bjc.6601682.
 56. Beckner ME, Gobbel GT, Abounader R, Burovic F, Agostino NR, *et al.* Glycolytic glioma cells with active glycogen synthase are sensitive to PTEN and inhibitors of PI3K and gluconeogenesis. *Lab Invest* 2005; 85(12): 1457–1470. doi: 10.1038/labinvest.3700355.
 57. Vignot S, Faivre S, Aguirre D, Raymond E. mTOR-targeted therapy of cancer with rapamycin derivatives. *Ann Oncol* 2005; 16(4): 525–537. doi: 10.1093/annonc/mdl113.
 58. Patel S, Doble B, Woodgett JR. Glycogen synthase kinase-3 in insulin and Wnt signalling: A double-edged sword? *Biochem Soc Trans* 2004; 32(5): 803–808. doi: 10.1042/BST0320803.
 59. Jiang R, Mircean C, Shmulevich I, Cogdell D, Jia Y, *et al.* Pathway alterations during glioma progression revealed by reverse phase protein lysate arrays. *Proteomics*

- 2006; 6(10): 2964–2971. doi: 10.1002/pmic.200500555.
60. Hutterer M, Gunsilius E, Stockhammer G. Molecular therapies for malignant glioma. *Wien Med Wochenschr* 2006; 156(11): 351–363. doi: 10.1007/s10354-006-0308-3.
 61. Grossman SA, Alavi JB, Supko JG, Carson KA, Priet R, *et al.* Efficacy and toxicity of the antisense oligonucleotide aprinocarsen directed against protein kinase C- α delivered as a 21-day continuous intravenous infusion in patients with recurrent high-grade astrocytomas. *Neuro Oncol* 2005; 7(1): 32–40. doi: 10.1215/S1152851703000353.
 62. Sachdev D, Yee D. Disrupting insulin-like growth factor signaling as a potential cancer therapy. *Mol Cancer Ther* 2007; 6(1): 1–12. doi: 10.1158/1535-7163.MCT-06-0080.
 63. Ardourel M, Blin M, Moret JL, Dufour T, Duc HT, *et al.* A new putative target for antisense gene therapy of glioma: Glycogen synthetase. *Cancer Biol Ther* 2007; 6(5): 719–723. doi: 10.4161/cbt.6.5.4232.
 64. Goudar RK, Shi Q, Hjelmeland MD, Keir ST, McLendon RE, *et al.* Combination therapy of inhibitors of epidermal growth factor receptor/vascular endothelial growth factor receptor 2 (AEE788) and the mammalian target of rapamycin (RAD001) offers improved glioblastoma tumor growth inhibition. *Mol Cancer Ther* 2005; 4(1): 101–112.
 65. Zhou X, Ren Y, Moore L, Mei M, You Y, *et al.* Downregulation of miR-21 inhibits EGFR pathway and suppresses the growth of human glioblastoma cells independent of PTEN status. *Lab Invest* 2010; 90: 144–155. doi: 10.1038/labinvest.2009.126.
 66. Premkumar DR, Arnold B, Jane EP, Pollack IF. Synergistic interaction between 17-AAG and phosphatidylinositol 3-kinase inhibition in human malignant glioma cells. *Mol Carcinog* 2006; 45(1): 47–59. doi: 10.1002/mc.20152.
 67. Downward J. Targeting RAS signalling pathways in cancer therapy. *Nat Rev Cancer* 2003; 3: 11–22. doi: 10.1038/nrc969.
 68. Sanson M, Laigle-Donadey F, Benouaich-Amiel A. Molecular changes in brain tumours: Prognostic and therapeutic impact. *Curr Opin Oncol* 2006; 18(6): 623–630. doi: 10.1097/01.cco.0000245322.11787.72.
 69. Ertl DA, Gleiss A, Sagmeister S, Haeusler G. Determining the normal range for IGF-I, IGFBP-3, and ALS: New reference data based on current internal standards. *Wien Med Wochenschr* 2014; 164 (17–18): 343–352. doi: 10.1007/s10354-014-0299-4.
 70. Trojan J, Briceno I. IGF-I antisense and triple-helix gene therapy of glioblastoma. In: A. Pantar (editor). *Evolution of the molecular biology of brain tumors and the therapeutic implications*. Vienna: InTech; 2013. p. 149–166.
 71. Vadgama JV, Wu Y, Datta G, Khan H, Chillar R. Plasma insulin-like growth factor-I and serum IGF-binding protein 3 can be associated with the progression of breast cancer, and predict the risk of recurrence and the probability of survival in African-American and Hispanic women. *Oncol* 1999; 57(4): 330–340. doi: 10.1159/000012052.
 72. Campbell MJ, Woodside JV, Secker-Walker J, Titcomb A, Leatham AJC. IGF status is altered by tamoxifen in patients with breast cancer. *Mol Pathol* 2001; 54(5): 307–310. doi: 10.1136/mp.54.5.307.
 73. Eppler E, Zapf J, Bailer N, Falkmer UG, Falkmer S, *et al.* IGF-I in human breast cancer: Low differentiation stage is associated with decreased IGF-I content. *Eur J Endocrinol* 2002; 146: 813–821. doi: 10.1530/eje.0.1460813.
 74. Wolk A, Anderson SO, Mantzoros CS, Trichopoulos D, Adami HO. Can measurements of IGF-I and IGFBP-3 improve the sensitivity of prostate cancer screening? *Lancet* 2000; 356(9245): 1902–1903. doi: 10.1016/S0140-6736(00)03266-9.
 75. Trojan J, Kopinski P, Drewa T, Powierska-Czarny J, Pacholska J, *et al.* Immunogenotherapy of prostate cancer. *Urol Pol* 2003; 56(2): 7–11.
 76. Mishra L, Bass B, Ooi BS, Sidawy A, Korman L, *et al.* Role of insulin-like growth factor-I (IGF-I) receptor, IGF-I, and IGF binding protein-2 in human colorectal cancers. *Growth Horm IGF Res* 1998; 8(6): 473–479. doi: 10.1016/S1096-6374(98)80300-6.
 77. Kopinski P, Ly A, Trojan J. Antisense therapies in oncology. In: A. Ly & D. Khayat (editors). *About cancer in Africa*. Paris: INCa; 2006. p. 659–671.
 78. Pitts TM, Tan AC, Kulikowski GN, Tentler JJ, Brown AM, *et al.* Development of an integrated genomic classifier for a novel agent in colorectal cancer: approach to individualized therapy in early development. *Clin Cancer Res*. 2010; 16(12): 3193–3204. doi: 10.1158/1078-0432.CCR-09-3191.
 79. Lee DY, Kim SJ, Lee YC. Serum insulin-like growth factor (IGF)-I and IGF-binding proteins in lung cancer patients. *J Korean Med Sci* 1999; 14(4): 401–404. doi: 10.3346/jkms.1999.14.4.401.
 80. Martenka J, Plato M, Kopyński P, Soja J, Szczeklik J, *et al.* (Polish) [Studies on insulin-like growth factor-I expression in human lower airways]. *Pol Merkur Lekarski* 2005; 19(113): 621–624.
 81. Upegui-Gonzalez LC, Trojan LA, Ly A, François J-C, Przewlocki R, *et al.* Antisense and triple-helix strategies in basic and clinical research: Challenge for gene therapy of tumors expressing IGF-I. In: D. LeRoith, W. Zumkeller, RC. Baxter (editors). *Insulin like-Growth Factor I*. New York: Landes Bioscience/ Eurekah & Kluwer Academic/Plenum Publishers; 2003. p. 357–366.
 82. Zumkeller W, Westphal M. The IGF/IGFBP system in CNS malignancy. *Mol Pathol* 2001; 54: 227–229. doi:10.1136/mp.54.4.227.
 83. Giovannucci E. Insulin-like growth factor-I and binding

- protein-3 and risk of cancer. *Horm Res Paediatr* 1999; 51(Suppl 3): 34–41. doi: 10.1159/000053160.
84. Yu H, Rohan T. Role of the insulin-like growth factor family in cancer development and progression. *J Natl Cancer Inst* 2000; 92(18): 1472–1489. doi: 10.1093/jnci/92.18.1472.
 85. Kopczak A, Stalla GK, Uhr M, Lucae S, Hennings J, *et al.* IGF-I in major depression and antidepressant treatment response. *Eur Neuropsychopharmacol* 2015; 25(6): doi: 10.1016/j.euroneuro.2014.12.013.
 86. Vicente M. *Avances en ingeniería genética* (Spanish). [Advances in genetic engineering]. Madrid: Consejo Superior de Investigaciones Científicas; 1994.
 87. Weinberg SL. Patient education as a part of critical care. *Heart Lung* 1974; 3(1): 47–48.
 88. Mayr E. Weismann and evolution. *J Hist Biol* 1985; 18(3): 295–329. doi: 10.1007/BF00138928.
 89. Yockey HP. Comments on “Let there be life; Thermodynamic reflections on biogenesis and evolution” by Avshalom C. Elitzur. *J Theor Biol* 1995; 176(3): 349–355. doi: 10.1006/jtbi.1995.0204.
 90. Darwin C. *The descent of man, and selection in relation to sex*. Rev. ed., New York: Merrill and Baker; 1874. doi: 10.5962/bhl.title.54341.
 91. Crick FH. The Croonian lecture, 1996: The genetic code. *Proc R Soc Lond B Biol Sci* 1967; 167(1009): 331–347.
 92. Depew DJ, Weber BH. *Evolution at a crossroads: The new biology and the new philosophy of science*. Cambridge MA, US: MIT Press; 1985.
 93. Monod J. *Chance and necessity: An essay on the natural philosophy of modern biology*. New York: Vintage Books; 1972.
 94. Rubenstein JL, Nicolas JF, Jacob F. Construction of a retrovirus capable of transducing and expressing genes in multipotential embryonic cells. *Proc Natl Acad Sci USA* 1984; 81(22): 7137–7140. doi: 10.1073/pnas.81.22.7137.
 95. Weintraub H, Izant JG, Harland RM. Anti-sense RNA as a molecular tool for genetic analysis. *Trends Genet* 1985; 1: 22–25. doi: 10.1016/0168-9525(85)90010-1.
 96. Hélène C. Control of oncogene expression by antisense nucleic acids. *Eur J Cancer* 1994; 30A(11): 1721–1726.
 97. Sharp PA. RNA interference—2001. *Genes Dev* 2001; 15: 485–490. doi: 10.1101/gad.880001.
 98. Anderson WF, Blaese RM, Culver K. Points to consider response with clinical protocol, July 6, 1990. *Hum Gene Ther* 2008; 1(3): 331–362. doi: 10.1089/hum.1990.1.3-331.
 99. Ferrari G, Rossini S, Nobili N, Maggioni D, Garofalo A, *et al.* Transfer of the ADA gene into human ADA-deficient T lymphocytes reconstitutes specific immune functions. *Blood* 1992; 80(5): 1120–1124.
 100. Pulkkanen KJ, Yla-Herttua S. Gene therapy for malignant glioma: Current clinical status. *Mol Ther* 2005; 12(4): 585–598.
 101. Johnson TR, Trojan J, Rudin SD, Ilan Ju, Tykocinski ML *et al.* Evoking an immune response to glioblastoma cells transfected with episome based plasmid expressing anti-sense transcripts to insulin like growth factor I. In: AJ. Levine & H.H. Schmidek (editors). *Molecular genetics of nervous system tumours*. New York: John Wiley & Sons; 1993. p. 387–400.
 102. Holtzman NA. Promoting safe and effective genetic tests in the United States: Work of the task force on genetic testing. *Clin Chem* 1999; 45(5): 732–738.
 103. Pergament E. New molecular techniques for chromosome analysis. *Best Pract Res Clin Obstet Gynaecol* 2000; 14(4): 677–690. doi: 10.1053/beog.1999.0104.
 104. Hoedemaekers R, Ten Have H. Genetic health and genetic disease. In: Launis V, Pietarinen J, Rääkkä J, (editors). *Genes and morality: New essays*. Ed. Amsterdam-Atlanta GA: Rodopi BV; 1999. p. 121–143.
 105. Ensenauer RE, Reinke SS, Ackerman MJ, Tester DJ, Whiteman DAH, *et al.* Primer on medical genomics Part VIII: Essentials of medical genetics for the practicing physician. *Mayo Clin Proc* 2003; 78(7): 846–857. doi: 10.4065/78.7.846.
 106. Wideroff L, Freedman AN, Olson L, Klabunde CN, Davis W, *et al.* Physician use of genetic testing for cancer susceptibility: Results of a national survey. *Cancer Epidemiol Biomarkers Prev* 2003; 12(4): 295–303.
 107. Snow K. The growing impact of genetics on health care: Do we have appropriate educational resources? *Mayo Clin Proc* 2001; 76(8): 769–771. doi: 10.1016/S0025-6196(11)63218-7.
 108. Pagon RA, Pinsky L, Beahler CC. Online medical genetics resources: A US perspective. *BMJ* 2001; 322: 1035–1037. doi: 10.1136/bmj.322.7293.1035.
 109. Glazier AM, Nadeau JH, Aitman TJ. Finding genes that underlie complex traits. *Science* 2002; 298(5602): 2345–2349. doi: 10.1126/science.1076641.
 110. Grady WM. Genetic testing for high risk colon cancer patients. *Gastroenterol* 2003; 124(6): 1574–1594. doi: 10.1016/S0016-5085(03)00376-7.
 111. Saleh N, Moutereau S, Azulay JP, Verny C, Simonin C, *et al.* High insulin like growth factor I is associated with cognitive decline in Huntington disease. *Neurology* 2010; 75 (1): 57–63. doi: 10.1212/WNL.0b013e3181e62076.
 112. Parente Pereira AC, McQuillin A, Puri V, Anjorin A, Bass N, *et al.* Genetic association and sequencing of the insulin-like growth factor 1 gene in bipolar affective disorder. *Am J Med Genet B Neuropsychiatr Genet* 2011; 156(2): 177–187. doi: 10.1002/ajmg.b.31153.
 113. Bondy CA, Werner H, Roberts CT Jr, LeRoith D. Cellular pattern of insulin-like growth factor-I (IGF-I) and type I IGF receptor gene expression in early organogenesis: Comparison with IGF-II gene expression. *Mol Endo-*

- crinol 1990; 4(9): 1386–1398. doi: 10.1210/mend-4-9-1386.
114. Love S, Louis DN, Ellison DW. Greenfield's neuropathology. 8th ed. Florida, USA: CRC Press; 2008. p. 521.
 115. Sturm MA, Conover CA, Pham H, Rosenfeld RG. Insulin-like growth factor receptors and binding protein in rat neuroblastoma cells. *Endocrinol* 1989; 124(1): 388–396. doi: 10.1210/endo-124-1-388.
 116. Trojan J, Johnson TR, Rudin SD, Blossey BK, Kelley KM, et al. Gene therapy of murine teratocarcinoma: Separate functions for insulin-like growth factors I and II in immunogenicity and differentiation. *Proc Natl Acad Sci USA* 1994; 91(13): 6088–6092. doi: 10.1073/pnas.91.13.6088.
 117. Trojan J. (French) [Expression of neuro-ectoblast in murine teratocarcinomas: Electron-microscopic and immunocytochemical studies, applications in embryology and in tumor pathology of central nervous system]. *Bull Inst Pasteur* 1984; 83: 335–385.
 118. Obrepalska-Stepelowska A, Kedzia A, Trojan J, Goździcka-Józefiak A. Analysis of coding and promoter sequences of the IGF-I gene in children with growth disorders presenting with normal level of growth hormone. *J Pediatr Endocrinol Metab* 2003; 16(9): 1267–1275. doi: 10.1515/JPEM.2003.16.9.1267.
 119. Zumkeller W. IGFs and IGF-binding proteins as diagnostic markers and biological modulators in brain tumors. *Expert Rev Mol Diagn* 2002; 2(5): 473–477. doi: 10.1586/14737159.2.5.473.
 120. Trojan J, Pan YX, Wei MX, Ly A, Shevelev A, et al. Methodology for anti-gene anti-IGF-I therapy of malignant tumours. *Chemother Res Pract* 2012; 2012(2012). doi: 10.1155/2012/721873.
 121. Sussenbach JS, Steenbergh PH, Holthuisen P. Structure and expression of the human insulin-like growth factor genes. *Growth Regul* 1992; 2(1): 1–9.
 122. Caliebe J, Broekman S, Boogaard M, Bosch CAJ, Ruivinkamp CAL, et al. IGF1, IGF1R and SHOX mutation analysis in short children born small for gestational age and short children with normal birth size (idiopathic short stature). *Horm Res Paediatr* 2012; 77(4): 250–260. doi: 10.1159/000338341.
 123. Wit JM. Diagnosis and management of disorders of IGF-I synthesis and action. *Pediatr Endocrinol Rev* 2011; 9(Suppl 1): 538–540.
 124. Metlapally R, Ki CS, Li YJ, Tran-Viet KN, Abbott D, et al. Genetic association of insulin-like growth factor-I polymorphisms with high-grade myopia in an international family cohort. *Invest Ophthalmol Vis Sci* 2010; 51(9): 4476–4479. doi: 10.1167/iavs.09-4912.
 125. Cao Y, Lindström S, Schumacher F, Stevens VL, Albanes D, et al. Insulin-like growth factor pathway genetic polymorphisms, circulating IGF1 and IGFBP3, and prostate cancer survival. *J Natl Cancer Inst* 2014; 106(6): dju085. doi: 10.1093/jnci/dju085.
 126. Gu F, Schumacher FR, Canzian F, Allen NE, Albanes D, et al. Eighteen insulin-like growth factor pathway genes, circulating levels of IGF-I and its binding protein, and risk of prostate and breast cancer. *Cancer Epidemiol Biomarkers Prev* 2010; 19(11): 2877–2887. doi: 10.1158/1055-9965.EPI-10-0507.
 127. Soerensen M, Dato S, Tan Q, Thinggaard M, Kleindorp R, et al. Human longevity and variation in GH/IGF-1/insulin signaling, DNA damage signaling and repair and pro/ antioxidant pathway genes: Cross sectional and longitudinal studies. *Exp Gerontol* 2012; 47(5): 379–387. doi: 10.1016/j.exger.2012.02.010.
 128. Gibbons JJ, Abraham RT, Yu K. Mammalian target of rapamycin: Discovery of rapamycin reveals a signaling pathway important for normal and cancer cell growth. *Semin Oncol* 2009; 36(Suppl 3): S3–S17. doi: 10.1053/j.seminoncol.2009.10.011.
 129. Piro A, Tagarelli G, Lagonia P, Quattrone A, Tagarelli A. Archibald Edward Garrod and alcaptonuria: “Inborn errors of metabolism” revisited. *Genet Med* 2010; 12: 475–476. doi: 10.1097/GIM.0b013e3181e68843.
 130. Koppenol WH, Bounds PL, Dang CV. Otto Warburg's contributions to current concepts of cancer metabolism. *Nat Rev Cancer* 2011; 11(5): 325–337. doi: 10.1038/nrc3038.
 131. Hanahan D, Weinberg RA. Hallmarks of cancer: The next generation. *Cell* 2011; 144(5): 646–674. doi: 10.1016/j.cell.2011.02.013.
 132. Parsons DW, Jones S, Zhang X, Lin JCH, Leary RJ, et al. An integrated genomic analysis of human glioblastoma multiforme. *Science* 2008; 321(5897): 1807–1812. doi: 10.1126/science.1164382.
 133. Yan H, Parsons DW, Jin G, McLendon R, Rasheed A, et al. *IDH1* and *IDH2* mutations in gliomas. *N Engl J Med* 2009; 360(8): 765–773. doi: 10.1056/NEJMoa0808710.
 134. Johnston Rohrbasser LB. Genetic testing of the short child. *Horm Res Paediatr* 2011; 76(Suppl 3): 13–16. doi: 10.1159/000330141.
 135. Backeljauw P, Bang P, Clayton PE, Geffner M, Woods KA. Diagnosis and management of primary insulin-like growth factor-I deficiency: Current perspectives and clinical update. *Pediatr Endocrinol Rev* 2010; 7(Suppl 1): 154–171.
 136. Hahn M, Saeger HD, Schackert HK. Hereditary colorectal cancer: Clinical consequences of predictive molecular testing. *Int J Colorectal Dis* 1999; 14(4): 184–193. doi: 10.1007/s003840050210.
 137. Weitzel JN. Genetic cancer risk assessment. *Cancer* 1999; 86(Suppl 11): 2483–2492. doi: 10.1002/(SICI)1097-0142(19991201)86:11+2483::AID-CNCR5>3.0.CO;2-4
 138. Hadley DW, Jenkins J, Dimond E, Nakahara K, Grogan L,

- et al.* Genetic counseling and testing in families with hereditary nonpolyposis colorectal cancer. *Arch Intern Med* 2003; 163(5): 573–582. doi: 10.1001/archinte.163.5.573.
139. Beauchamp TL, Childress JF. *Principles of biomedical ethics*. 4th ed., New York: Oxford University Press; 1994.
140. Pellegrino ED. Ethics. *JAMA* 1995; 273(21): 1674–1676. doi: 10.1001/jama.1995.03520450044022.
141. Gould SJ. Message from a mouse: It takes more than genes to make a smart rodent, or high-IQ humans. *Time* 1999; 154(11): 42.
142. Tang TLP, Weatherford EJ. Perception of enhancing self-worth through service: The development of a Service Ethic Scale. *J Soc Psychol* 1998; 138(6): 734–743. doi: 10.1080/00224549809603258.
143. Egan MF, Goldberg TE, Kolachana BS, Callicott JH, Mazzanti CM, *et al.* Effect of COMT Val^{108/158} Met genotype on frontal lobe function and risk of schizophrenia. *Proc Natl Acad Sci USA* 2001; 98(12): 6917–6922. doi: 10.1073/pnas.111134598.
144. Hamer DH. A linkage between DNA markers on the X chromosome and male sexual orientation. *Science* 1993; 261(5119): 321–327. doi: 10.1126/science.8332896.
145. Nelkin D, Andrews LB. Whose genes are they anyway? *Chron High Educ* 1999; 45: B6.
146. Lippman A. Prenatal genetic testing and screening: Constructing needs and reinforcing inequities. *Am J Law Med* 1991; 17(1–2): 15–50.
147. Lippman A. Led (astray) by genetic maps: The cartography of the human genome and health care. *Soc Sci Med* 1992; 35(12): 1469–1476. doi: 10.1016/0277-9536(92)90049-V.
148. Rothman BK. *Genetic maps and human imaginations: The limits of science in understanding who we are*. New York: WW Norton and Company; 1998.
149. Lupton D. *Medicine as culture: Illness, disease and the body in Western societies, Part 2*. London: Sage Publications Ltd; 1994.
150. Levin DM, Solomon GF. The discursive formation of the body in the history of medicine. *J Med Philos* 1990; 15(5): 515–537. doi: 10.1093/jmp/15.5.515.
151. Temple LKF, McLeod RS, Gallinger S, Wright JG. Essays on science and society: Defining disease in the genomics era. *Science* 2001; 293(5531): 807–808. doi: 10.1126/science.1062938.
152. Green ED, Watson JD, Collins FS. Human Genome Project: Twenty-five years of big biology. *Nature* 2015; 526(7571): 29–31. doi: 10.1038/526029a.